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Additions to the liverwort flora of the Kermadec Islands, New Zealand Botanical Region

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Abstract

Five liverwort species are reported for the first time from the New Zealand Botanical Region, all from Raoul Island in the Kermadec Islands group: *Ceratolejeunea belangeriana*, *Notoscyphus lutescens* and *Lepidolejeunea integristipula* (new records for their respective genera in New Zealand), *Acrolejeunea pycnoclada* var *pycnoclada* and *Cheilolejeunea ceylanica*. Keys to Australasian *Acrolejeunea* and vitate species of *Cheilolejeunea* subg. *Xenolejeunea* are provided.

Introduction

The Kermadec Islands group $(29^{\circ} 15' - 31^{\circ} 30' \text{ S}, 177^{\circ} 55' - 179^{\circ} 00' \text{ W})$ (Fig. 1) is the northernmost extension of the New Zealand Botanical Region (sensu Allan 1961; Wardle 1991). Of the 29 or so islands, islets, rock stacks and emergent reefs that make up the group, Raoul Island at 2943 ha and elevations up to 518 m. a.s.l. is the largest, highest and only island supporting a dense cover of indigenous forest. Although the vascular flora of the Kermadec Islands are especially well documented (e.g. Sykes 1977; Sykes & West 1996) the bryophyte flora (particularly the hornwort and liverwort flora) remains poorly known. Campbell in Sykes (1977), based largely on gatherings from Raoul and Macauley Islands, listed 20 hornwort and liverwort taxa, and Beever et al. (1996) accepted 52 mosses for the island group. Since then there have been only a few publications dealing with new bryophyte records for the island (Campbell 1997; Fife & de Lange 2009; Renner & de Lange 2011; Renner et al. 2013a), new endemic taxa (Engel & Merrill 1999; Renner 2005) and/or reinstatements of synonymised species (So 2001; Engel & Merrill 2010). Currently, there is one liverwort endemic to the Kermadec Islands, *Plagiochila pacifica* Mitt. which is known only from Raoul Island, (So 2001; Engel & Merrill 2010). A second species, Radula erigens (M.A.M.Renner & Braggins) M.A.M.Renner (Renner 2005; Renner et al. 2013b), described initially as an endemic subspecies of R. cordiloba Taylor (Renner 2005), is now regarded as synonymous with R. javanica Gottsche (Renner 2014). There are no known endemic mosses (de Lange & Beever *in press*) or hornworts occurring in the Kermadec Islands.

In May 2011, the Auckland War Memorial Museum (AK) led the *Kermadec Biodiscovery Expedition* visiting all the main islands, islets and rock stacks, except for Curtis, Haszard, 'Haszardette', Milne and Dougal Rocks. As a result, five taxa new to the Kermadec Islands, as well as for the New Zealand Botanical Region, were identified:

Acrolejeunea pycnoclada (Taylor) Schiffner var. pycnoclada, Ceratolejeunea belangeriana (Gottsche) Steph., Cheilolejeunea ceylanica (Gottsche) R.M.Schust. & Kachroo, Lepidolejeunea integristipula (Jack & Steph.) R.M. Schust., and Notoscyphus lutescens (Lehm. & Lindenb.) Mitt. A description of the habitat of each of these taxa and their conservation status in the New Zealand Botanical Region are provided using the New Zealand Threat Classification System (Townsend et al. 2008).

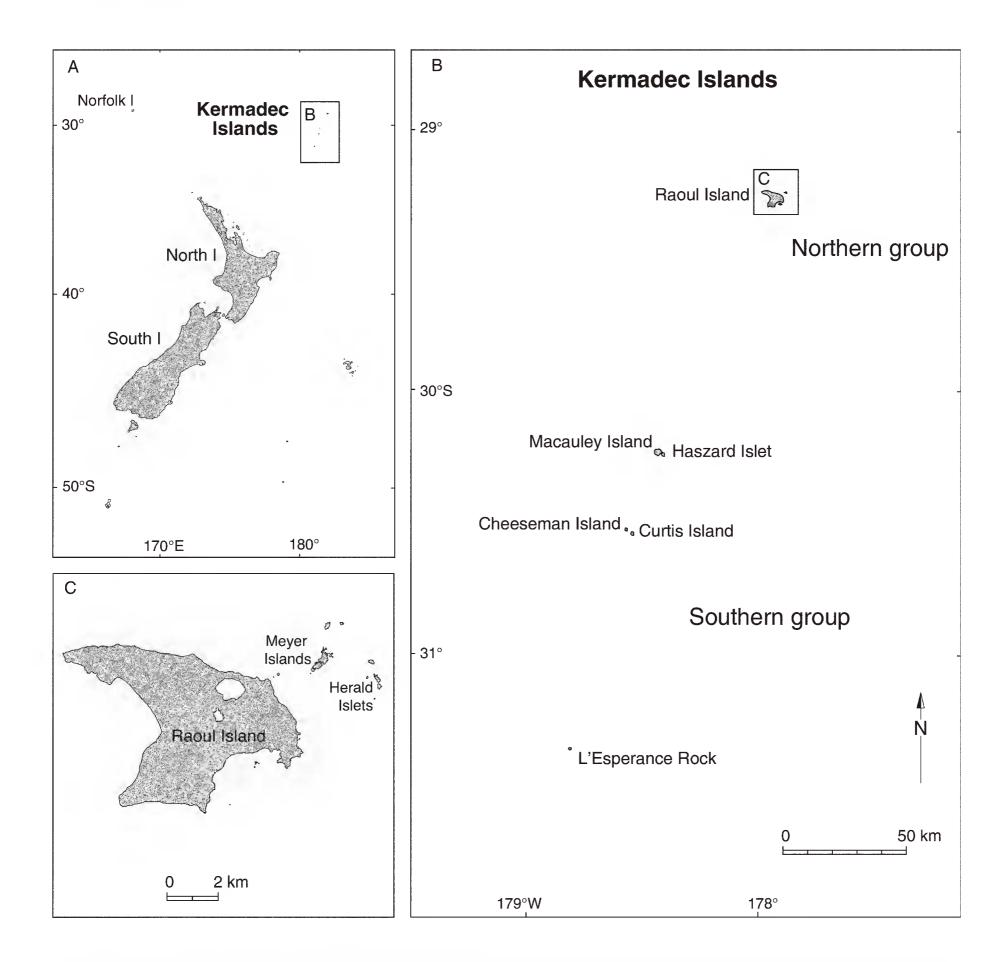


Fig. 1. Location and spatial extent of the Kermadec Islands. A, General map showing position of the Kermadec Islands relative to Norfolk Island and the North and South Islands of New Zealand; B, Details of Kermadec Islands; C, Herald Islets, Meyer Islands, Raoul Island of the Kermadec Islands.

New records for the New Zealand Botanical Region

Acrolejeunea pycnoclada (Taylor) Schiffner var. pycnoclada: Engl. & Prantl, Natürlichen Pflanzenfamilien I, 3(1): 128 (1893).

Ptycanthus pycnocladus Taylor Journal of Botany 5: 835 (1846).

Lejeunea pycnocladus (Taylor) Mitt. Journal of the Proceedings of the Linnean Society, Botany 5: 111 (1861). Ptychocoleus pycnocladus (Taylor) Steph. Species Hepaticarum 5: 52 (1912).

Type: MALAYSIA. Malacca, *Cantor s.n.* (holotype fide Gradstein, 1975: FH! Taylor Herbarium; isotypes: G! MANCH, NY, S, U, W).

Corticolous. Plant olive-green. Stems to at least 8 mm long, mostly shorter, appressed to bark. 0.3 mm (dry)–0.9 mm (wet) wide; 90–110 µm diameter, ventral merophyte of 4 or 5 cells in width, Cortical cells subrectangular to irregular, 20–30 × 12–22 µm with c. 17 cortical cells surrounding c. 20 medullary cells. Branches mostly of *Lejeunea* type but occasionally *Frullania* type. Leaves imbricate when dry, spreading but not squarrose when wet. Lobe c. 540×330 µm, broad, shortly crescent-shaped with apex rounded, incurved. Dorsal base not auriculate. Ventral margin short and \pm recurved. Keel curved, not decurrent and cells not or weakly protuberant. Median leaf cells ovoid, 22×15 –16 µm, base cells larger 29–31 × 16–17 µm, trigones medium-sized, Y-shaped (sides swollen), intermediate thickenings scattered to scarce. Oil-bodies not present. Lobule narrowly ovate-rectangular with apex slightly oblique, free margin almost straight. Teeth (2 or)3, each 1 or 2 cells long, erect, first cell at tip of free margin, hyaline papilla at base of second tooth (the 'apical tooth'), lobule is about half length of lobe. Underleaf, slightly overlapping base of adjacent underleaf, broadly oval, c. 190 µm long, c. 320 wide. Median cells ovoid, 15– 18×13 –15 µm, slightly larger at base, 22– $25 \times (9$ –)10(–13) µm. Rhizoid disc small. Sporophyte not seen. Fig. 2

Specimen examined: New Zealand, Kermadec Islands Nature Reserve, Raoul Island, Sunshine Valley, near Sunshine Bivy, 16 May 2011, *P.J. de Lange K688* (AK325478, F, NSW).

Recognition: The female bracts of *Acrolejeunea pycnoclada* are distinctive in being cucullate, and closely adpressed to one another. The lobule bears two small triangular teeth that are two cells broad at the base, and in this character differ from other Australian species. The lobule of *A. securifolia* has 3 or 4, 1–3-celled teeth that are usually clawed toward and behind the ventral surface of the lobule, and so are difficult to observe. The lobules of *A. mollis* Steph. have a single prominent, triangular tooth. *Acrolejeunea pycnoclada* from Raoul Island does not have the distinctly punctate surface ornamentation found in Australian plants (Renner 2013).

Distribution: Acrolejeunea pycnoclada var. pycnoclada is widespread through Africa, Indo-Malesia and the Pacific (Gradstein 1975), whereas in the New Zealand Botanical Region it is known only from the single gathering from Raoul Island in the Kermadec Islands.

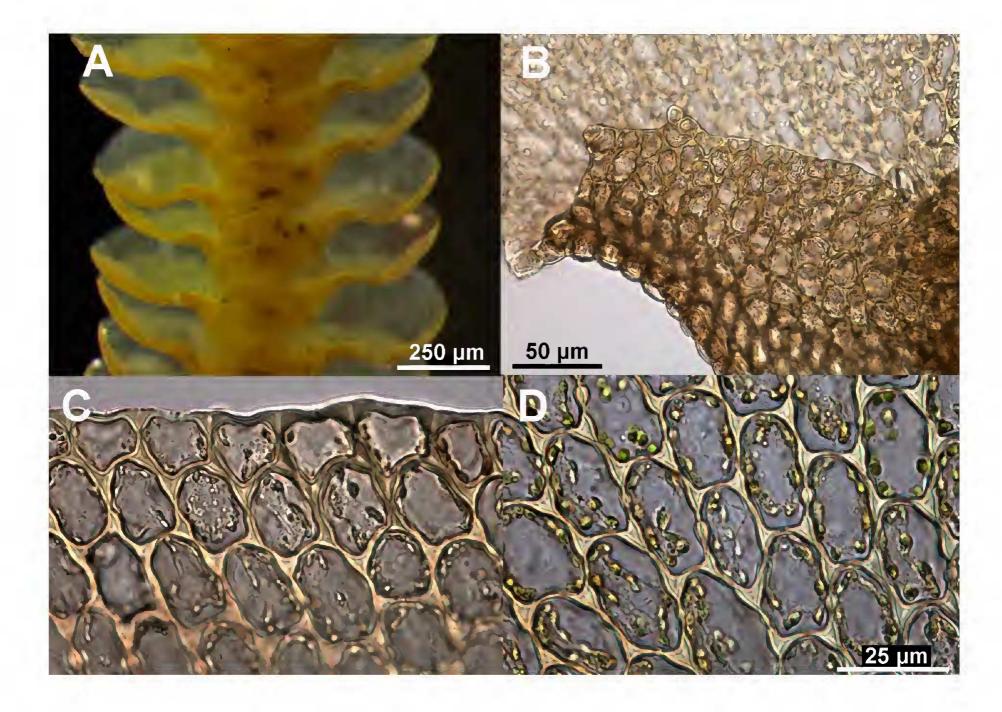


Fig. 2. *Acrolejeunea pycnoclada* var. *pycnoclada* A: ventral view of shoot. B: lobule showing teeth. C: marginal leaf-lobe cells. D: medial leaf-lobe cells. All from AK325478.

Habitat: The gathering came from the branch of a *Metrosideros kermadecensis* W.R.B.Oliv. *Acrolejeunea pycnoclada* var. *pycnoclada* grew in association with the liverwort *Frullania rostrata* (Hook.f. & Taylor) Hook.f. & Taylor, moss *Calymperes tenerum* Müll.Hal., ferns *Pyrrosia eleagnifolia* (Bory) Hovenkamp and *Asplenium* aff. *haurakiense* (Brownsey) Ogle, and a number of lichens including *Heterodermia obscurata* (Nyl.) Trevis., *Hypotrachyna costaricensis* (Nyl.) Hale, *Parmotrema reticulatum* (Taylor) M.Choisy, and *Pseudocyphellaria argyracea* (Delise) Vain.

Conservation Status: Although *A. pycnoclada* var. *pycnoclada* is widespread outside of New Zealand, it is known only from a single gathering from Raoul Island in the New Zealand Botanical Region, where it was abundant on *Metrosideros kermadecensis* in that locality. Therefore, the conservation status of this species is here regarded as Data Deficient, qualified OL [One Location] and SO [Secure Overseas].

Key to Australasian Acrolejeunea

A key to all *Acrolejeunea* species currently known from Australia and New Zealand that builds on the key to Australian species in Renner (2013) is presented below.

Plants large, shoots to 1.5 mm wide, reddish brown; leaf lobe cells arranged in radiating rows; lobules narrowly pyriform, keel straight most its length, inclined at 30-45° to stem (measured on shoot apex side Plants medium to small, shoots 0.8–1.3 mm wide, mid-green, yellow-green or grey-green, never reddish; leaf lobe cells not arranged in radiating rows; lobules narrowly obpyriform to trapeziform, keel straight or Dioicous; androecia intercalary on leading shoots; female bracts in two gyres, bract lobe apices obtuseacute, bract lobes particularly those of inner gyre often reflexed; perianths exserted above bracts at maturity, underleaves ovoid to longitudinally oblong; lobule second tooth shorter or longer than lobule Autoicous; androecia on short lateral branches; female bracts in one gyre, bract lobe apex rounded, bract lobes not reflexed; perianths immersed within bracts at maturity; underleaves longitudinally oblong c. 0.2× the shoot width; lobule second tooth longer than lobule first tooth 3 3 Perianth not or only slightly emergent, plicate 1/4–1/3 of length, beak 7-8 cells long, underleaf 0.5 mm wide Perianth emergent for ½ its length, plicate over ½-2/3 its length, beak 3-4 cells long, Underleaf 0.6-0.8 mm Lobule with four teeth, the first of one or two cells, the remainder unicellular, evenly distributed along the broadly rounded lobule free antical margin; underleaf weakly obovate; female bract margins entire Lobule with three teeth, variously one to three celled, unevenly distributed along the lobule free antical Lobule arch one or two cells, (first lobule tooth one or two cells from the junction of the lobe and lobule), first and/or second lobule tooth hooked, lobule carinal region broadly pyriform, keel curved; leaf lobe cells finely punctulate; female bracts cucullate, margins entire; perianth rostrum 8–12 cell tiers high Lobule arch 4 or 5 cells (first lobule tooth four or five cells from the junction of the lobe and lobule), lobule teeth not hooked, lobule carinal region narrowly pyriform, keel straight; leaf lobe cells unornamented; female bracts plane, margins scalloped; perianth rostrum 3–7 cell tiers high.....

Ceratolejeunea belangeriana (Gottsche) Steph. Species Hepaticarum 5: 396 (1913).

Lejeunea belangeriana Gottsche in Gottsche et al. Synopsis Hepaticarum 398 (1845).

Type: REÚNION. Without more specific locality, Belanger ex herb. Steph. 'Original' (BM!).

Ceratolejeunea oceanica (Mitt.) Steph. in Reinecke Botanische Jahrbücher 23: 310 (1897).

Lejeunea oceanica Mitt. Flora Vitiensis 414 (1871 [1873]).

Ceratolejeunea oceanica (Steph.) Horik. Acta Phytotaxonomica et Geobotanica 13: 214. (1943) nom. illeg.

Lopholejeunea oceanica Steph. Species Hepaticarum 6: 378 (1923).

Type: SAMOA. Without more specific locality, T. Powell s.n. (NY).

Ceratolejeunea renistipula Steph. Botanische Jahrbücher 20: 236 (1895).

Type: TAHITI. Without more specific locality, M. Vesco 85 (G-11910!).

Forming loose patch of dispersed shoots mixed with and growing on other bryophytes. Shoots goldenbrown to black (in dried material), medium-sized for genus, to 10 mm long and 0.8–1.3 mm wide, branching irregular, shoot system monomorphic. Stems with seven inflated cortical cells and up to 20 smaller medulla cells, cortical and medulla cell walls golden-brown to dark-brown pigmented, walls not heavily thickened, but with triangular trigones at cell junctions. Dorsal leaf-free strip absent. Leaf-lobes rotund to asymmetrically ovate with apex near postical lobe margin, 480–750 µm long, 380–500 µm wide, apex rounded on smaller to acute on larger lobes, margin with scattered triangular teeth comprised of 3–5 cells around apex, concave, imbricate, interior margin extending over stem to opposite stem margin and obscuring stem in dorsal view. Leaf lobes not vittate, but consistently with 2 or 3 scattered ocelli at leaf base, medial cells rounded to oblong, 20–28 µm long and wide, walls with continuous secondary thickening more pronounced at junctions forming triangular trigones, medial thickenings occasional but obscure, cells becoming larger toward leaf-lobe base, marginal cells quadrate to oblong, 8–15 µm long and wide, walls appearing to lack secondary thickening and occasionally also pigmentation in the free external wall. Oil-bodies not known. Lobules dimorphic, often explanate, when developed 120–150 µm long and 90–120 µm wide at base, approximately one twentieth the lobe area, pyriform, keel curved, carinal region strongly and continuously inflated along entire keel length and lobule width, lobule antical margin inrolled, not visible in ventral view, lobule arch 3 cells, lobe-lobule junction lying postical to the base of the first lobule tooth, first lobule tooth 1-celled, triangular with a sharp apex, long axis perpendicular to stem so as to point sideways and extend beyond the lobule exterior margin, lobule second tooth unicellular, more or less incorporated into lobule margin, lobule papilla attached to lobule margin immediately above the first tooth. Underleaves rotund, cordate, contiguous to imbricate, broadest near base, 300–500 μm long, 500–750 μm wide, bifid to 0.3×, sinus V-shaped, narrow, c. 70 μm deep, lobes acute, with a single cell at lobe apex. Underleaf insertion U-shaped across two ventral cortical cells, lateral basal cells of underleaf not inflated, underleaf base not cordate. Oil-bodies not known. Asexual reproduction absent. Paroicous. Androecia on modified, determinate, secondary shoots bearing 2–8 pairs of bracts. Gynoecia terminal on primary or secondary shoots, subtended by 2 subfloral innovations when located on primary shoots and 1 subfloral innovation when on secondary shoots, subfloral innovations always with Pycnolejeuneoid merophyte sequence, female bracts elliptic-ovate, acute, denticulate, bracteole narrowly elliptic, lobes acute, divided to 0.5×, margin irregular to denticulate, perianths to 1 mm long, more or less immersed among bracts, 5-carinate, lateral and ventral carinae cornute so perianth bears 4 horns that project above bracts, stipa not developed in observed perianths. Sporophyte not seen. Fig. 3.

Specimen examined: New Zealand, Kermadec Ecological Region and District, Kermadec Islands Nature Reserve, Raoul Island, base of, Sunshine Valley Waterfall, 29° 17′ 10.8′′ S 177° 55′ 45.7′′ W, c. 53 m, 16 May 2011, *P.J. de Lange K1147* (AK329755).

Recognition: *Ceratolejeunea belangeriana* is recognised by its dark brown to black colouration in combination with large, bifid underleaves whose base is cordate, the obscurely dentate leaf-lobe margin, and presence of ocelli at leaf-lobe base. This species is similar to many of the larger species of *Lopholejeunea* in its golden brown to black colour, but easily distinguished by its bifid underleaves. When fertile, the horn-bearing perianths are also characteristic of this species.

Distribution: Ceratolejeunea belangeriana is widely distributed throughout the Pacific and, in its current broadly accepted circumscription, extends as far west as South Africa.

Habitat: *Ceratolejeunea belangeriana* was collected at the base of Sunshine Valley Waterfall, where it grew as scattered shoots admixed with *Plagiochila stephensoniana* Mitt. among a large mat of *Radula javanica* that was carpeting a damp boulder directly below the ephemeral waterfall.

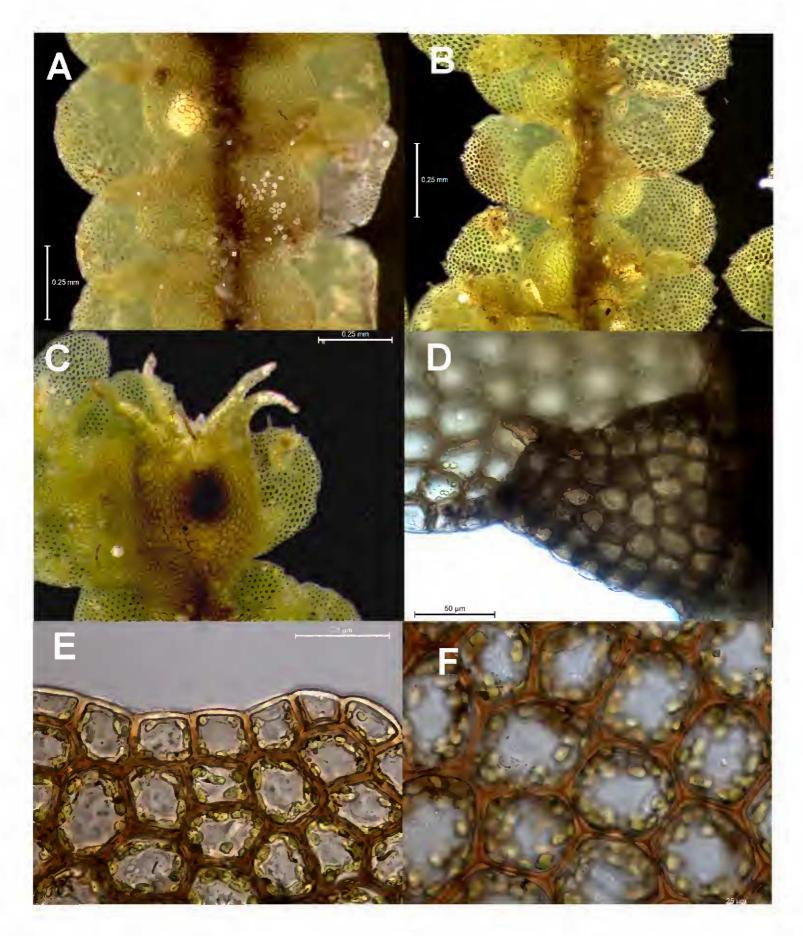


Fig. 3. *Ceratolejeunea belangeriana* A: ventral view of large shoot. B: ventral view of small shoot. C: Dorsal view of perianth. D: ventral view of lobule. E: marginal leaf-lobe cells. F: medial leaf-lobe cells. All from AK329755.

Conservation status: *Ceratolejeunea belangeriana* is known from the New Zealand Botanical Region from the single Raoul Island gathering. We assign this species a conservation status of Data Deficient, qualified OL and SO, for the New Zealand part of its range.

Cheilolejeunea ceylanica (Gottsche) R.M.Schust. & Kachroo Journal of the Linnean Society, Botany 56: 509 (1961).

Pycnolejeunea ceylanica (Gottsche) Schiffn. Hepaticae ... aus Engler-Prantl, Die naturlichen Pflanzenfamilien : 124 (1893).

Pycnolejeunea ceylanica (Gottsche) Steph. Hedwigia 28: 259 (1889), nom. inval.

Lejeunea ceylanica Gottsche Synopsis Hepaticarum 3: 359 (1845).

Type: CEYLON. "inter Rad. boryanum. mis. Hooker" (isotype: G-19377).

Cheilolejeunea verdoornii (G.Hoffm.) R.M.Schust. & Kachroo. Journal of the Linnean Society, Botany 56: 509 (1961).

Pycnolejeunea verdoornii G.Hoffm. Annales Bryologici 8: 118 (1935).

Type: INDONESIA. "Java orient., Res. Pasoewean, in mm. Tengger, in silvis primigen, in decliv. G. Ajek-Ajek," ca 2100 m, *anno* 1930, *Verdoorn s.n.* (holotype: JE, isotype: FH) syn. fide Thiers (1992).

Euosmolejeunea setchelii Pearson. Publications of the Carnegie Institution of Washington 20: 148 (1924).

Type: SAMOA. Tutuila Is., Aua-Aofono trail, Jun–Jul 1920, Setchell 1161a/1277a (holotype: MANCH) syn fide Thiers (1992).

Lejeunea connivens Gottsche ex Schiffn. Forschungsreise S.M.S. "Gazelle". 4, Botanik: 32. (1889 [1890]).

Type: INDONESIA. Moluccas, Amboina Is. "in litore boreali-occidentali ...", leg. *Naumann*, 7 Jun.1875. syn fide Soderström (unpublished data).

Pycnolejeunea arietina Tixier. Bulletin de la Société d'Histoire Naturelle de l'Afrique du Nord 63: 9 (1972).

Type: MALAYSIA. Kedah distr, Kedah Peak, 900 m, leg. Tixier 6069 (holotype: PC slide) syn. fide Grolle (1979).

Forming loose patches of dispersed shoots mixed with and growing on other bryophytes on tree trunks. Shoots brown-green (when fresh), light-brown (when dried), medium-sized for genus, to 10 mm long, 0.9-1.3 mm wide, branching infrequent and irregular, shoot system monomorphic. Stems with seven cortical cells and up to 15 smaller medulla cells, cortical and medulla cell walls with yellow-brown pigmentation, external free cortical cell wall, radial longitudinal cortical walls and inner tangential cortical walls continuously thickened, with bulging trigones at cell junctions, medulla cell walls with bulging trigones confluent across medial walls, constricting the cell lumen. Dorsal leaf-free strip present, dorsal cortical cell row not or only partly covered by leaf insertion lines. Lobes elliptic-oblong, slightly falcate and then almost reniform, 440-750 µm long, 360–430 µm wide, flat, imbricate, interior margin extending over stem to opposite stem margin and obscuring stem in dorsal view, lobe apex broadly rounded. Leaf lobes vittate, basal and medial cells oblong, 15–25 µm long, 10–20 µm wide, with concave trigones, medial thickenings common but obscure, transition to marginal cells gradual on all sides of vitta, marginal cells quadrate to oblong, 8–10 µm long and wide, walls continuously thickened, triangular trigones present. Oil-bodies not known. Lobules monomorphic, always well developed, 200–250 μm long, 65–115 μm wide, large lobules one seventh the lobe area, rectangular to falcate, keel straight to arched, carinal region weakly and continuously inflated along keel length, lobule antical margin not inrolled, visible in ventral view, lobule arch arched, 1–3 cells, lobe-lobule junction lying postical to base of first lobule tooth, first lobule tooth 1-celled, partially or completely incorporated into the exterior lobule margin, lobule second tooth multicellular, 3-5 cells, 2 cells wide at base, uniseriate for 2 or 3 cells, originating perpendicular to shoot axis, but curved backward, apex pointing toward postical lobe margin, overtopping first tooth, lobule papilla attached to lobule margin immediately above first tooth. Underleaves elliptic-rotund, remote, to contiguous, broadest at midpoint, 260–340 μm long, 220–280 μm wide, bifid to 0.4× on larger underleaves, sinus V-shaped, narrow, 70–120 µm deep, lobes obtuse, with a single cell at lobe apex. Underleaf insertion transverse across two ventral cortical cells, lateral basal cells of underleaf weakly inflated, underleaf base not cordate. Oil-bodies not known. Asexual reproduction not known. Dioicous (?). Androecia terminal on primary shoots or lateral on secondary branches. Gynoecia, perianths and sporophytes not seen. Fig. 4.

Specimens examined: New Zealand, Kermadec Islands Nature Reserve, Kermadec Islands northern group, Raoul Island, Smiths Bluff track just past Mahoe, 29° 17' 24" S 177° 56' 59" W, 330 m, 15 May 2011, *P.J. de Lange K731* (AK325641, NSW900223); Moumoukai-Prospect track (roughly half-way between Moumoukai and Prospect), 29° 16' 30" S 177° 55' 17" W, 15 May 2011, *P.J. de Lange K733* (AK325644, NSW851682). In both specimens *C. ceylanica* is scattered over and among *Radula javanica*.

Recognition: *Cheilolejeunea ceylanica* is readily recognised by the combination of long rectangular lobules with a more or less straight keel extending about two-thirds of the leaf-lobe width, the second lobule tooth 3–6 cells long (terminology follows Renner 2012), and the group of enlarged cells, 4 or 5 cells long, 3 or 4 cells wide, in the leaf-lobe base that are weakly differentiated from surrounding cells. The shape of the lobule places the species into subg. *Xenolejeunea*.

Distribution: Cheilolejeunea ceylanica has a paleotropical distribution, and has been reported from Japan, Thailand, the Philippines, and Indonesia (Mizutani 1980), the wet tropics of Queensland (Thiers 1992), and Fiji (Pócs et al. 2011). The record for the Kermadec Islands extends the distribution south and east, and, on the basis of this record, its occurrence throughout the Pacific should be anticipated.

Habitat: *Cheilolejeunea ceylanica* is an epiphyte that can inhabit all tree surfaces. It may form near-pure patches or occur in multi-species mats with other bryophytes. On Raoul Island *Cheilolejeunea ceylanica* is known from two gatherings in which it grew scattered with *Lejeunea gracilipes* and *Lepidolejeunea integristipula* as an epiphyte (or epipmuscicole) on the liverwort *Radula javanica*.

Conservation Status: *Cheilolejeunea ceylanica* is known from the New Zealand Botanical Region by two gatherings made on Raoul Island. We assign this species a conservation status of Data Deficient, with qualifiers OL and SO.

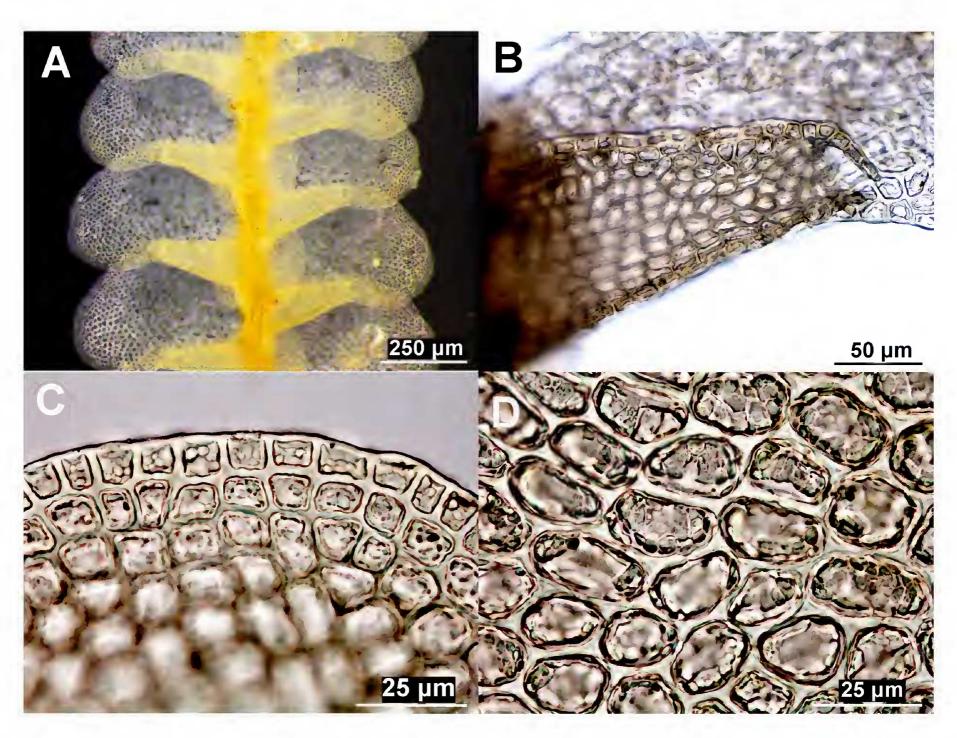


Fig. 4. *Cheilolejeunea ceylanica* A: ventral view of shoot. B: moniliform second lobule tooth pointing toward postical lobe margin. C: marginal leaf-lobe cells. D: basal leaf-lobe cells. All from AK325644.

Key to vittate and ocellate species of Cheilolejeunea subg. Xenolejeunea

The key to vittate and ocellate Australasian species of *Cheilolejeunea* subg. *Xenolejeunea* from Renner (2012) is reproduced below.

- 2 Vitta cells markedly differentiated from surrounding cells, vitta zone up to 15 cells long and 10 cells wide 3
- Vitta cells weakly differentiated from surrounding cells, vitta zone 4 or 5 cells long, 3 or 4 cells wide

 Cheilolejeunea ceylanica (Gottsche) R.M.Schust. & Kachroo
- Vitta cells oblong, without distinct medial wall thickenings, vitta zone extending to two thirds the lobule length, second lobule tooth on largest lobules up to 3 cells long, lobule fused to lobe across 1 or 2 cells, explanate lobules with two teeth, cladia present

Lepidolejeunea integristipula (Jack & Steph.) R.M.Schust. Phytologia 45: 425 (1980).

Pycnolejeunea integristipula Jack & Steph. Beihefte zum Botanischen Centralblatt 60: 107 (1894).

Type: FIJI. Viti insulae, Ovalau, in monte Tana-lailai, anno 1864, *Graeffe* (holotype *fide* Piippo (1986): G-20388).

Known from a few shoots growing scattered through bryophytes epiphytic on *Radula javanica*, yellow-brown when dry, medium-sized for genus, to 15 mm long, 0.8–0.9 mm wide, branching infrequent and irregular, shoot system monomorphic. Stems with external and internal walls unthickened except for weak concave trigones of primary wall around cell wall junctions, 7 cortical cells and 5–10 smaller medullary cells. Dorsal leaf-free strip absent. Lobes rotund, 350–400 µm long and wide, plane. Lobe apex rounded, postical margin straight toward keel, antical margin continuously curved, exceeding opposite stem margin. Basal and medial cells irregularly hexagonal, 15–25 µm long, 9–18 µm wide, thin-walled with weak concave trigones, medial thickenings absent, marginal cells quadrate, 12–18 µm long and wide, thin-walled, external wall protruding, triangular in outline, lobe margin serrulate as a result. Lobules relatively tiny, 70–80 µm long and wide, about one thirtieth the lobe area, almost sphaeroidal with inflation of carinal region, keel curved, mostly in outer half, base perpendicular to stem. Carinal region inflated, lobule antical margin inrolled, obscured in ventral view except for area in vicinity of first tooth; lobule arch straight, 2 cells, angle between arch and keel 10° at lobelobule juncture, lobe-lobule junction lying slightly postical to the base of the first lobule tooth. First lobule tooth 1-celled, triangular, oriented perpendicular to shoot axis or pointing outward. Lobule second tooth not or only weakly differentiated from adjacent marginal cells. Underleaves reniform, remote to contiguous, broadest at midpoint, 250–310 µm long, 520–600 µm wide, entire. Underleaf insertion transverse across 2 ventral cortical cells, lateral basal cells of underleaf inflated, underleaf base not cordate. Oil-bodies not known. Asexual reproduction absent. Sporophyte not seen. Fig. 5.

Specimen examined: New Zealand, Kermadec Islands Nature Reserve, Kermadec Islands northern group, Raoul Island, Moumoukai-Prospect track (c. half-way between Moumoukai and Prospect, 29° 16' 30" S 177° 55' 17" W, 15 May 2011, *P.J. de Lange K733* (AK325644, NSW851682). Scattered over *Radula javanica* with *Cheilolejeunea ceylanica*.

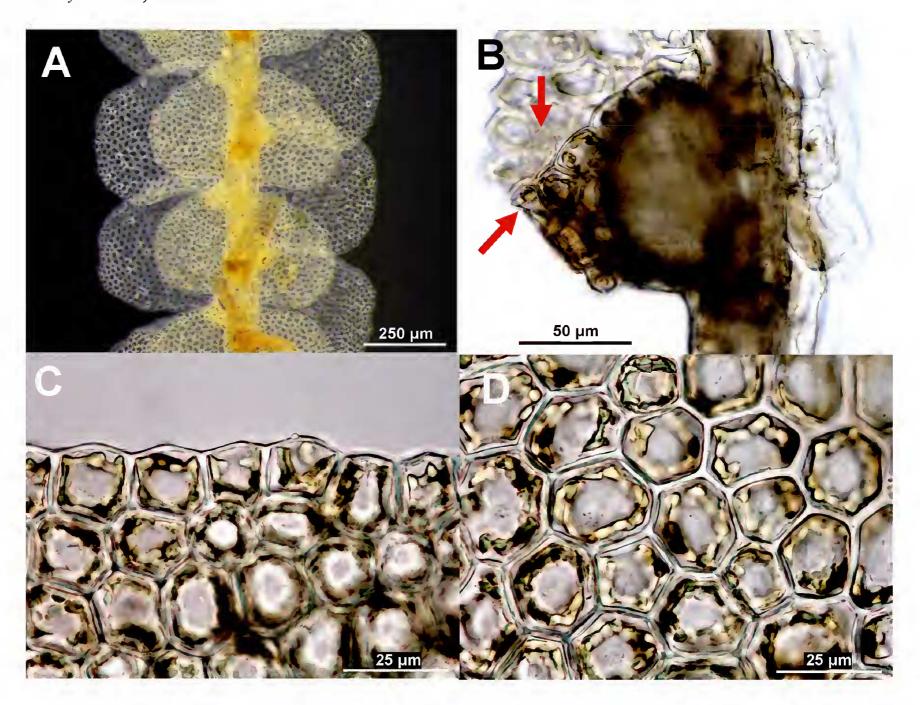


Fig. 5. *Lepidolejeunea integristipula* A: ventral view of shoot. B: lobule showing lobule papilla (indicated by upper arrow) and sideways pointing, triangular first lobule tooth (indicated by lower arrow). C: marginal leaf-lobe cells. D: basal leaf-lobe cells. All from AK325644.

Recognition: Among species of Lejeuneaceae currently known from New Zealand, *Lepidolejeunea integristipula* is recognisable by the isolobous shoot systems having large, reniform, entire underleaves; the lobules with a prominent sideways pointing first lobule tooth; and the leaf-lobe cell walls lacking secondary pigmentation.

Entire underleaves are more commonly associated with species belonging to subfamily Ptychanthoideae, such as *Archilejeunea*, *Lopholejeunea* and *Thysananthus* in New Zealand. *Lepidolejeunea integristipula* can be separated from all New Zealand species of subfam. Ptychanthoideae by its lack of secondary pigmentation in the leaf-cell walls and by the crenulate-serrulate leaf-lobe margins where each minute tooth is formed by angular projecting cell lumen. The New Zealand plants of *L. integristipula* do not have obvious ocelli and, therefore, bear resemblence to *Lejeunea mimula*, a species distributed from Malaysia to Fiji. However, several micro-morphological characters separate these two species. In *Lejeunea mimula*: 1) the first lobule tooth is forward-pointing, and situated half-way between the lobe-lobule and the lobule-stem junction; 2) the leaf-lobe margin of *L. mimula* is entire; 3) the cell walls have discrete medial thickenings; and 4) the leaf-lobe cell surface bears punctate to finely granular ornamentation. In contrast in *Lepidolejeunea integristipula*: 1) the first lobule tooth is sideways pointing and is situated over the lobe-lobule junction; 2) the leaf-lobe margin is crenulate-serrulate; 3) the cell walls lack medial thickenings; 4) the leaf-lobe cell surface is smooth.

Distribution: *Lepidolejeunea integristipula* is distributed throughout the Asian-Oceanic regions from Borneo to Fiji and south to the Wet Tropics Bioregion of north-eastern Queensland (Australia) (Pócs et al. 2011; Pócs and Streimann 2006). The Kermadec Islands record represents a new global southern limit for this species.

Habitat: Throughout its wide geographic distribution *Lepidolejeunea integristipula* is typically a lowland plant, inhabiting a range of microsites within lowland mesophyll rainforest, including the surfaces of rotting logs, trunks of rainforest trees, and leaves of large-leaved trees and shrubs. It may form pure mats. When growing on branches, twigs and leaves, the species may become semi-pendulous and form near-pure wefts. On Raoul Island *L. integristipula* grows as scattered stems among a mat of larger bryophyte species dominated by *Radula javanica* and including *Cheilolejeunea ceylanica* and *Lejeunea gracilipes*.

Conservation status: *Lepidolejeunea integristipula* is known from the New Zealand Botanical Region by one gathering made on Raoul Island. We assign this species a conservation status Data Deficient, with qualifiers OL and SO.

Notoscyphus lutescens (Lehm. & Lindenb.) Mitt. Flora Vitiensis 407 (1871 [1973]).

Acolea lutescens (Lehm. & Lindenb.) Trevis. Memorie del Reale Istituto Lombardode Scienze e Lettere. Serie 3, Classe di Scienze Matematiche e Naturali 4: 395 (1877).

Cesius lutescens (Lehm. & Lindenb.) Kuntze. Revisio Generum Plantarum 2: 834 (1891).

Gymnanthe lutescens (Lehm. & Lindenb.) Mitt. Journal of the Linnean Society, Botany 5: 100 (1861).

Gymnomitrion lutescens (Lehm. & Lindenb.) Gottsche Synopsis Hepaticarum 1: 4 (1844).

Nardia lutescens (Lehm. & Lindenb.) Steph. Bulletin de l'Herbier Boissier, séries 1, 5: 81 (1897).

Jungermannia lutescens Lehm. & Lindenb. Novarum et Minus Cognitarum Stirpium Pugillus 4: 16 (1832).

Type: INDIA. Habitat in India orientali (BM, G, S, W).

Note: heterotypic synonyms not listed here.

Terricolous, forming a light green loose weft with individual shoots to $14 \times 1.2-1.4$ mm, anisophyllous. Branching infrequent, ventral-intercalary, stems slender, terete, $94-120~\mu m$ diameter, 7 cells high in transverse section; cortical cells thick walled in surface view variable in size and shape $37-59\times 19-26~\mu m$; rhizoids $9-15~\mu m$ diam produced in groups at or near underleaf bases, occasionally scattered on stem. Leaves unlobed or slightly bilobed, alternate, succubous, nearly horizontal, flat when damp, dorsally assurgent when dry, contiguous, shortly oblong, $430-700\times 270-500~\mu m$, margin entire; not inserted to stem midline dorsally leaving one row of cortical cells leaf-free, dorsal bases slightly decurrent; leaf cells are variable in size the median cells $(31-)51-55\times 24-26(-31)~\mu m$, basal cells to $40-47~\mu m$ wide, trigones triangular, bulging slightly Y-shaped $9\times 7.5~\mu m$, no intermediate thickening; cell surface with c. 20 low striae over entire surface. Underleaves small, bifid, with lobes lying parallel to stem, unequal one being shorter by 1 or 2 cells, $250-374\times 94-187~\mu m$, with longest lobes $150-210~\mu m$ long, with 4-7 uniseriate cells based on 1 biseriate row; 1 or 2(or~3) single-celled (rarely 2-celled) lateral spurs variably developed on the disc, or lateral spur rarely absent; cell walls are thick (c. $4.4~\mu m$) and swollen at corners, cells rectangular, length: width ratio 1.3-1.4:1, tip cell being narrower (length to width ratio c. 1.5:1). Oil bodies 2-4(-5) per cell ovoid to spherical, homogeneous, pale yellow-brown, variable in size, $9-13~\mu m$ diam. Sporophyte not seen. Fig. 6.

Specimen examined: New Zealand, Kermadec Islands Nature Reserve, Kermadec Islands northern group, Raoul Island, Prospect – Mahoe Hut Track (near Prospect Peak, eastern flank), 15 May 2011, *P.J. de Lange K730* (AK325635, CHR, F).

Recognition: The underleaves are either as wide as the stem or narrower than stem, with long setaceous lobes orientated parallel to stem, with 1 small tooth between base of lobe and insertion on each disc margin. The leaf-lobe is ornamented by up to c. 20, long, low striae that are orientated perpendicular to stem, on dorsal leaf-lobe surface only. Neither the underleaves nor the striate leaf surface ornamentation occur in combination in other phenetically similar genera, including *Chilocyphus* and *Saccogynidium*.

Remarks: Since the description of oil-bodies was taken from dried material, some degeneration may have occurred.

Distribution: *Notoscyphus lutescens* has been reported from South Africa, Madagascar, India, China, Japan, Philippines, Indonesia, Papua New Guinea, north-eastern Australia (Queensland), as well as New Caledonia, Hawai'i, Fiji and Samoa (Schuster 2002). Its presence on the Kermadec Islands is considered as being within the distribution given by Schuster (2002).

Habitat: *Notoscyphus* was collected from the south-eastern flank of Prospect Peak, growing in deep shade at the back of 'tree cave' made by a *Metrosideros kermadecensis*. There it grew by itself on damp peaty soil and grit festooned amongst *Metrosideros* and *Cyathea milnei* Hook. roots.

Conservation status: *Notoscyphus lutescens* is known from the New Zealand Botanical Region by a single gathering made on Raoul Island. We assign this species a conservation status of Data Deficient, with qualifiers OL and SO.

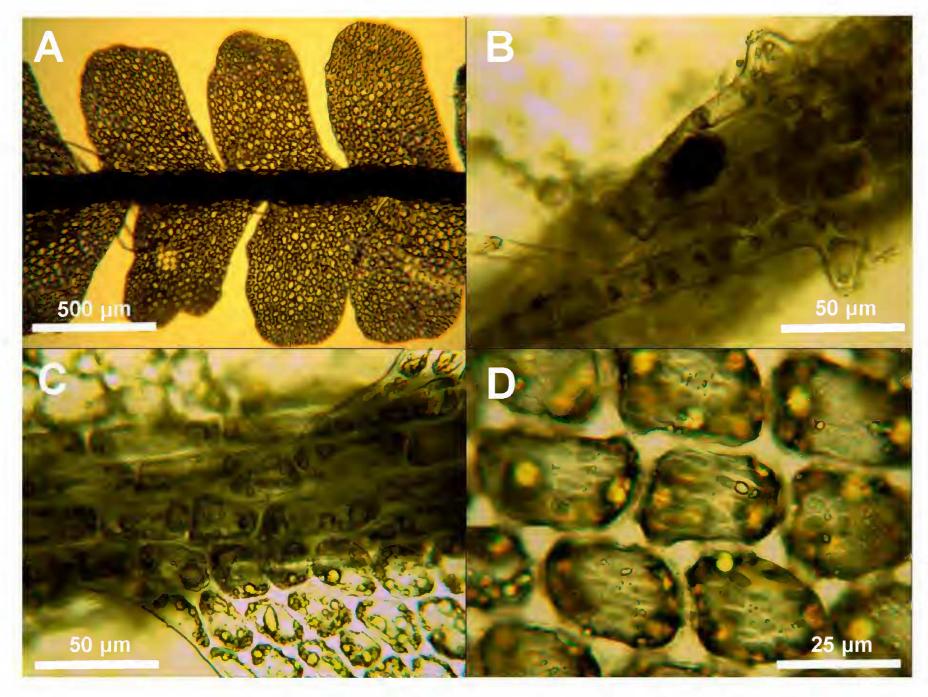


Fig. 6. *Notoscyphus lutescens* A: ventral view of shoot. B: underleaf. C: dorsal stem surface. D: leaf cell surface. All from AK325635.

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References

- Allan HH (1961) Flora of New Zealand. Vol. I. (Government Printer, Wellington)
- Beever JJ, Fife AJ, West CJ (1996) Mosses of the Kermadec Islands, northern New Zealand. *New Zealand Journal of Botany* 34: 463–471. http://dx.doi.org/10.1080/0028825X.1996.10410127
- Campbell EO (1977) Hepaticopsida (Hepaticae) and Anthocerotopsida In: Sykes WR (ed) *Kermadec Islands Flora an annotated checklist*. Wellington, *New Zealand Department of Scientific and Industrial Research Bulletin* 219: 200–205.
- Campbell EO (1997) Some additions and alterations to the liverwort and hornwort flora of the Kermadec Islands. *New Zealand Botanical Society* 49: 8–10.
- de Lange PJ, Beever JE. (In Press) A checklist of the mosses of the Kermadec Islands group. *Auckland Museum Bulletin* 20: c. 40 pp.
- Engel JJ, Smith-Merrill GL (2010) Studies on New Zealand Hepaticae. 39–55. More new taxa, combinations, typifications and synonymy in *Plagiochila* from New Zealand (Plagiochilaceae). *Nova Hedwigia* 91: 501–517. http://dx.doi.org/10.1127/0029-5035/2010/0091-0501
- Fife AJ, de Lange PJ (2009) *Calymperes tenerum* Müll.Hal. (Calymperaceae) on the Chatham Islands, New Zealand. *Australasian Bryological Newsletter* 57: 14–16.
- Gradstein SR (1975) A taxonomic monograph of the genus *Acrolejeunea* (Hepaticae) with an arrangement of the genera of Ptychanthoideae. *Bryophytorum Bibliotheca* 4: 1–182, 24 Plates.
- Mizutani M (1980) Notes on the Lejeuneaceae. 3. Some Asiatic species of the genus *Cheilolejeunea*. *Journal of the Hattori Botanical Laboratory* 47: 319–331.
- Piippo S (1986) A monograph of the genera *Lepidolejeunea* and *Luteolejeunea* (Lejeuneaceae, Hepaticae). *Acta Botanica Fennica* 132: 1–69.
- Pócs T, Streimann H (2006) Contributions to the bryoflora of Australia, I. *Tropical Bryology* 27: 19–24.
- Pócs T, Sass-Gyarmati A, Naikatini A, Braggins JE, von Konrat MJ (2011) New liverwort (Marchantiophyta) records for the Fiji Islands. *Telopea* 13: 455–494.
- Renner MAM (2005) Additions to the *Radula* (Radulaceae: Hepaticae) Floras of New Zealand and Tasmania. *Journal of the Hattori Botanical Laboratory* 97: 39–79.
- Renner MAM (2012) Further insight into lobule teeth homology in Lejeuneaceae subf. Lejeuneoideae from *Cheilolejeunea oscilla*, a new species from Australia. *The Bryologist* 115: 536–556.
- Renner MAM (2013) A new subspecies of *Acrolejeunea arcuata* and notes on other Australasian Lejeuneaceae. *Phytotaxa* 83: 39–53.
- Renner MAM (2014) *Radula* subg. *Radula* in Australasia and the Pacific (Jungermanniopsida). *Telopea* 17: 107–167. http://dx.doi.org/10.7751/telopea20147553
- Renner MAM, de Lange PJ (2011) Additions to the Lejeuneaceae flora of New Zealand: new species from the Kermadec Islands and range extensions of New Zealand species into the South Pacific. *New Zealand Journal of Botany* 49: 421–433.
- Renner MAM, Devos N, Brown EA, von Konrat MJ (2013a) A revision of Australian species of *Radula* subg. *Odontoradula*. *Australian Systematic Botany* 26: 408–447. http://dx.doi.org/10.1071/SB13035
- Renner MAM, Söderstöm L, Hagborg A, von Konrat MJ (2013b) Notes on early land plants today. 29. A new combination in *Radula* (Radulaceae, Marchantiophyta). *Phytotaxa* 81: 12–14. http://dx.doi.org/10.11646/phytotaxa.81.1.5
- Schuster RM (2002) Austral Hepaticae part II. Nova Hedwigia Beiheft 119:1-606.
- So ML (2001) Studies on *Plagiochila* subgenus *Metaplagiochila* (Plagiochilaceae, Hepaticae). *Australian Systematic Botany* 14: 677–688. http://dx.doi.org/10.1071/SB00033
- Sykes WR (1977) Kermadec Islands Flora an annotated checklist. Wellington, New Zealand Department of Scientific and Industrial Research Bulletin 219: 1–216.
- Sykes WR, West CJ (1996) New records and other information on the vascular flora of the Kermadec Islands. *New Zealand Journal of Botany* 34: 447–462. http://dx.doi.org/10.1080/0028825X.1996.10410126
- Thiers BM (1992) A re-evaluation of *Cheilolejeunea* subg. *Xenolejeunea*. *Tropical Bryology* 5: 10–21.
- Townsend AJ, de Lange PJ, Duffy CAJ, Miskelly CM, Molloy J, Norton DA (2008) New Zealand threat classification system manual. (Department of Conservation, Wellington)
- Wardle P (1991) Vegetation of New Zealand. (Cambridge University Press, Cambridge)